

The Risk Of Several Fungi Associated With Bird Waste

Zaidan Kalaif Imran¹ , Rusul Issam Ali²

Correspondence Author: Zaidan Kalaif Imran

Email:zaidan_omran@yahoo.com

ABSTRACT: In Babylon province and rounded cities , samples of dried waste were obtained and studied for the presence of fungi of potentially pathogenic nature. There was a high proportion of *Candida* spp , *Cryptococcus neoformans* and *Rhodotorula mucilaginosa* and other fungi obtained from the dry droppings. The aim of the present study was to survey filamentous fungi and yeasts associated with dropping of domestic birds in the Babylon province and rounded cities. During 2013–2014, 191 samples of dropping west were collected cultured and examined by dilution plate method .Filamentous fungi and yeasts colonies growths appearing on the Sabouraud's Dextrose Agar (SDA) medium were microscopically examined and the cultures obtained were identified on the basis of their microscopic and colonial morphology. A total of 2506 fungal isolates (included 2104 yeast isolates and 402 filamentous isolates were collected, *Candida* spp. constituted the majority (38.22 % of the 2104 isolates, represented by mainly *C.albicans* and *C.Krusei*, *Geotrichum* spp. *Blastomyces* spp., *Macrorhabdus* and *Cryptococcus*. These yeasts had proteolytic activities .conclusion indicated that *Blastomyces* spp. *Macrorhabdus* and *Cryptococcus* were recorded for the first time associated with bird waste in Iraq. *Cryptococcus* had Phospholipase activities .waste of birds is a finding be sources of risk on the public health.

Keywords: Bird ,dropping ,filamentous fungi ,yeasts, Phospholipase , .Iraq.

H. capsulatum to avian habits has been reviewed by Ajello (1964).and the organism has been reported in droppings from chickens, pigeons, and bats (Lehan and Furcolow, 1957; Stoker, 1964). Most Candidiasis and Cryptococcosis infections are mild and occur without symptoms(Lazera et al., 1996). Diffuse pulmonary infection is often asymptomatic and unrecognized. Persons with weakened immune systems, however, are more susceptible to symptomatic infection. (Ellis et al., 1990,).Fungal flora was representative the possible sources of infection to man (Mwaba et al, 2001) .

Although the possibility of their droppings having been collected is small,Feathers and hairs are a rich source of keratin and are regarded as one of the reservoirs of keratin-degrading microorganisms. Birds carry keratinophilic fungi passively through the intact feather (Mandeel and

1.Introduction

Large populations of roosting birds may present a disease sources, the most serious health risks from contact or inhaling of the fungal spores from disease organisms that grow in the nutrient-rich accumulations of bird droppings and dispersal , feathers and debris under a roost the association between dried droppings and the isolation of fungi was first described by Emmons (1955). Birds and their droppings can carry over 60 other diseases many of them are airborne and can be transferred to humans just by being around droppings(Littman and Borok, 1968; Zarrin et al., 2010). Most of early studies on the fungi associated with bird dropping were performed before 1970s .These studies reported that yeast has been isolated from pigeon droppings in London by Randhawa, Clayton, and Riddell (1965) and by Partridge and Winner (1965). The relationship of

recovery of keratinophilic fungi with the addition of hair-bait. Hence the latter technique was employed for all the samples of faeces. The plates were incubated at 30°C. filamentous fungi and yeasts were identified based on Wattab 2002. CHROMagar candida was used as confirmatory tests for *Candida* spp. (Nadeem et al 2010) Yeast were cultured on egg yolk agar plates for detection Phospholipase activities.

2.2. Isolation and identification of dropping fungi

Loop full of dropping suspension transferred to plates of Sabouraud dextrose agar (Hi Media, Mumbai, India) supplemented with chloramphenicol (Falcon Chemox Pvt, Ltd, Ghaziabad, India) at a concentration of 0.05 mg/ml. The cultures were microscopically examined to check for purity and sub-cultured to get pure cultures. The purified fungal isolates were identified up to species level as far as possible by a detailed study of their colonial characters and microscopic morphology in lactophenol blue mounts, and comparison of their characteristics with the descriptions of the species in the standard books and manuals (10–12). The study was approved by the local Ethics Committee.

Frequency % = $\frac{\text{No. of observation in which colony appear}}{\text{total number of observation recorded}} \times 100$ (Adhikari et al., 2004)

Occurrence % = $\frac{\text{No. of fungus in which sample appear}}{\text{total number of samples}} \times 100$ (Maria and Sridhar, 2003)

2.3. Determination of phospholipase activity

The phospholipase activity of *C. albicans* and *Cryptococcus* was detected by the method of (Samaranayake et al, 1984). Approximately 5 µL of standard inoculum of test strain containing 10⁸ *Candida* cells/mL was aseptically inoculated onto egg yolk agar. The plates were dried at room temperature and then incubated at 37°C for 48 h.

(Mancianti 2001). Some of these fungi are species of non-dermatophytes or dermatophytes known to cause cutaneous infections of keratinized tissues, viz skin, feather, hair and nail (dermatophytoses) of humans and animals (Efuntoye and Fashanu, 2002). Several workers have investigated the occurrence of Candidiasis fungi, *Coccidioides* and other fungi on a wide variety of birds in different countries as in Euripi, Africa, Austere, Filipich and Parker, 1993; Multu et al., 1997; Schulze and Heidrich 2001; Hanka et al., 2010; Martins et al., 2006; Phalen et al., 2007; Jansson et al., 2008;. Unfortunately, no previous studies about fungi associated with birds dropping were performed in Iraq. The aim of the present study was to survey filamentous fungi and yeast associated with dropping of domestic birds in the Babylon province and rounded cities.

2. Material and Methods

2.1. Collection and processing of samples of bird dropping

Samples collected from around Egg chickens, Meat chickens fields and cages of domestic birds. The dropping were collected from 8 different species of birds: Egg chickens, Meat chickens, *Sirrus leucopygius*, *Sernius canaria*, *Melopsittacus undulates*, *Nymphicus hollandicus*, *Aggornis* spp. and Pigeon. Throughout period extended from October 2013-April 2014 dry droppings from Babylon and rounded cities, 191 samples of dropping originated from were collected in plastic pages. 0.5 g of the dropping samples were diluted in sterile water 0.5 ml of suspension were powdered into Sabouraud's Dextrose Agar medium with 0.05 g/l chloramphenicol. The samples were placed in clean plastic zip bags which were labelled and transported promptly to the laboratory. The plates containing the samples of dropping were incubated at room temperature (22–30 °C) for 5-7 depending on the rate of growth, and periodically moistened with small quantities of SDW. Initially some samples of feathers were processed on soil plates. A comparison of the results showed better

<i>Rhizomucor</i>	20	28	0.25	1.11
<i>Cladosporium</i>	1	4	10.471	0.2
<i>Aspergillus flavus</i>	8	10	4.18	0.4
<i>A. terreus</i>	7	11	3.66	0.43
<i>A. fumigatus</i>	10	53	5.23	2.11
<i>A. niger</i>	29	39	15.18	1.6
<i>A. candidus</i>	5	9	2.61	0.35
<i>Cryptococcus</i> sp.	5	5	2.617	0.2
<i>Candida</i> spp.	73	710	38.2	28.32
<i>Macrorhabdus</i>	8	900	4.189	35.9
<i>Blastomyces</i>	2	2	1.04	0.079
<i>Geotrichum</i>	17	497	8.90	19.82
المجموع	191	2506		~100%

The plates were examined for the presence of precipitation zone around the colony. The presence of precipitation zone indicated expression of phospholipase enzyme.

3.Results :

Out of 191 samples of dropping waste of 8 species of birds examined, 2506 isolates of filamentous fungi and yeasts were recovered. The distribution of the isolates according to the species of birds investigated is more with Egg chickens, Meat chickens, , *Sernius canaria* and *Sirrus leucopygius*. *Aspergillus* spp., a well-known genus of filamentous fungi, constituted the majority (4.5%) followed by *Penicillium* spp (2.9%) of the 2506 isolates. Regarding the relative distribution of different species of filamentous fungi, . *Macrorhabdu* was the commonest species (35.9%) followed by *Geotrichum* and *Candida* spp (19.82,28.3%) respectively (Table 1). *Candida* spp showed different colors on CHROMagar (Figure 1). and *Cryptococcus* were showed phospholipase activity (Figure 2).

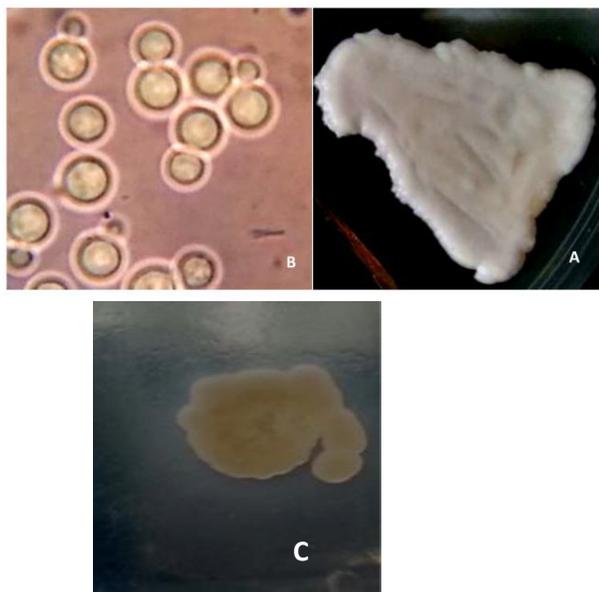


Figure 2: *Cryptococcus* sp isolated from bird dropping A-colony, B-Capsule of yeast, C- Phospholipase production.

Other species of fungi recovered from waste in the present study included 92 isolates

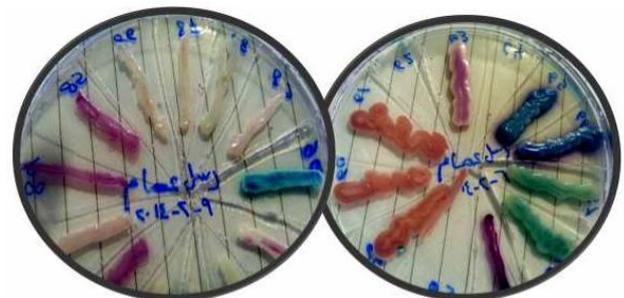


Figure 1: Colonies color of *Candida* spp and *Cryptococcus* sp isolated from bird dropping on CHROMagar.

Table 1: Fungi associated with bird dropping

Fungi	Samp les No.	Colo nies No.	% Occurr ence	% freque ncy
<i>Rhizopus</i>	44	92	23.036	3.66
<i>Penicilliu m</i>	17	72	8.90	2.87
<i>Fusarium</i>	2	2	1.047	0.1
<i>Mucor</i>	31	82	16.23	3.27

Ajello, L. (1964). Relationship of *Histoplasma capsulatum* to avian habitats. Pubi. Hlth Rep. (Wash.), 79: 266-270.

Denton, J. F., and Salvo, A. F. di (1968). The prevalence of *Cryptococcus neoformans* in various natural habitats. Sabouraudia, 6,213-217.

Efuntoye MO, Fashanu SO. (2002).Occurrence of keratinophilic fungi and dermatophytes on domestic birds in Nigeria. Mycopathologia, 153: 2987–9.

Ellis, D. H., and Pfeiffer, T. J. (1990). Natural habitat of *Cryptococcus neoformans* var. *gattii*. *J. Clin. Microbiol.* 28: 1642–1644.

Emmons, C. W. (1955). Saprophytic sources of *Cryptococcus neoformans* associated with the pigeon (*Columba li via*). *Amer. J.Hyg.*, 62:227-232.

Filippich L.J and Perry R.A.(1993). Drug trials against Megabacteria in birds in budgerigars (*Melopsittacus undulates*). *Australian Veterinary Practitioner.*23:184-189.

Hanka K, Koehler K, Kaleta E.F, Sommer D and Burkhardt E. (2010).*Macrorhabdus ornithogaster*: detection in companion birds, poultry and pigeons, morphological characterization and examination of *in vitro* cultivation. *Praktische Tierarzt.*91:390-395.

Jansson DS, Brojer C, Mattsson R, Feinstein R, Morner T and Segerstad CH. (2008). Mycotic proventriculitis in gray partridges (*Perdix perdix*) on two game bird farms. *Journal Zoo Wildlife Medicine.*39:428-437.

Kawakita, S., and Uden, N. van. (1965). Occurrence and population densities of yeast

of *Rhizopus*,(28) *Rhizomucor* (2),*Fusarium* and five isolates of *Cryptococcus* sp., Table1.

4.DISCUSSION

The present report on the occurrence of yeasts and filamentous fungi on dropping of birds in Babylon province and rounded cities constitutes the first study of its kind in Iraq. The predominance of *Geotrichum* spp ,*Candida* spp.is in agreement with that reported in surveys of bird dropping associated yeasts and filamentous fungi in several other countries (Filiphich and Parker, 1993 ;Multu *et al.*, 1997; Schulze and Heidrich 2001;; Phalen *et al.*, 2007;Martins *et al.*, 2006; Jansson *et al.*, 2008; Hanka *et al.*,2010). *Macrorhabdus*. *Geotrichum* and *C.albicans* were the most frequent species, followed by mucorales fungi, these results coincidence with Rad (2013). Phospholipases facilitate the invasion of the host mucosal epithelia by hydrolyzing one or more ester linkages in glycerophospholipids Singhai *et al.* ,2012 . In our study phospholipase was the major virulent factor expressed by *Cryptococcus* isolates . Screening of phospholipase production in *Cryptococcus* isolates can be used as an important parameter to differentiate invasive strains from noninvasive colonizers. our conclusion referred to presence of many of pathogenic and opportunistic filamentous and yeasts representative sources of risk fungal disease on the human health.

ACKNOWLEDGEMENTS

We wish to thank All women Science college for help in the processing of soils samples, and also to our colleagues in Laboratory of techniques supply of equipment and culture media. and all workers of chicken fields and bird cages .

References

Adhikari, A., Sen, M. M., Gupta - Bhattacharya, S. and Chanda, S. (2004).Airborne viable, non-viable and allergenic fungi in a rural agricultural area of India: a 2- year study at five outdoor sampling stations. *Sci. Total Environ.* 326: 123 - 141.

patients treated under local conditions. *Postgrad. Med. J.* 77: 769–773.

Nadeem S.G., Hakim S.T., Kazm S.U. (2010). Use chromoagar candida medium for the presumptive identification of *Candida* species directly from clinical specimens in resource-limited setting. *Libyan J Med* 5:1-6.

Partridge, B. M., and Winner, H. I. (1965). *Cryptococcus neoformans* in bird droppings in London. *Lancet*, 1: 1060-1061.

Rad, F. (2013). Isolation of *Cryptococcus neoformans* from pigeon excreta in Qazvin. *Life Science Journal*. 10(1).

Randhawa, H. S., Clayton, Y. M., and Riddell, R. W. (1965). Isolation of *Cryptococcus neoformans* from pigeon habitats in London. *Nature (Lond.)*, 208, 801.

Samaranayake, L. P., Raeside, J. M. & MacFarlane, T. W. (1984). Factors affecting the phospholipase activity of *Candida* species in vitro. *J Med Vet Mycol* 22: 201–207.

Schulze, C. and Heidrich, R. (2001). Megabakterien-assoziierte proventridulitis beim nutzgefugel in Brandenburg. *Dtsch Tierarztl Wochenschr* 108: 264–266.

Stoker, D. J. (1964). Histoplasmosis in Cyprus: Report of two cases. *Brit. med. J.*, 11: 793-795.

Zarrin, M., Jorfi, M., Amirrajab, N., and Rostami, M. (2010). Isolation of *Cryptococcus neoformans* from pigeon droppings in Ahwaz, Iran; 40 (2): 313-316.

species in the digestive tracts of gulls and terns. *J. gen. Microbiol.*, 39: 125-129.

Lazera, M. S., Salmito Cavalcanti, M. A., Londero, A. T., Trilles, L., Nishikawa, M. M., and Wanke, B. (2000). Possible primary ecological niche of *Cryptococcus neoformans*. *Med. Mycol.* 38: 379–383.

Lehan, P. H., and Furcolow, M. L. (1967). Epidemic histoplasmosis. *J. chron. Dis.* 5: 489-503.

Littman M.L, Borok R (1968). Relation of the pigeon to Cryptococcosis: natural carrier state, heat resistance and survival of *Cryptococcus neoformans*. *Mycopathol Mycol Appl.* 35:922-933.

Mandeeel Q, Mancianti F. (2011). Keratinophilic fungi on feathers of common clinically healthy birds in Bahrain. *Mycoses.* 54: 71–77.

Maria, G.L. and Sridhar, K.R. (2003). Diversity of filamentous fungi on woody litter of five mangrove plant species from the south west of India. *Fungal Diversity* 14:109-126.

Martins N.R.S, Horta A.C, Siqueira A.M, Lopes S.Q, Resende J.S, Jorge M.A, Assis R.A, Martins N.E, Fernandes A.A, Barrios P.R, Costa T.J.R and Guimaraes L.M.C. (2006). *Macrorhabdus ornithogaster* in ostrich, rhea, canary, zebra finch, free range chicken, turkey, guinea-fowl, columbina pigeon, toucan, chucker partridge and experimental infection in chicken, Japanese quail and mice. *Arquivo Brasileiro de Medicina Veterinaria Zootecnia*. 58:291-298.

Mutlu, O.F., Seckin, S., Ravelhofer Hildebrand RA and Grimm F. (1997). Proventriculus in fowl caused by megabacteria. *Tierarztl Pratische*. 25:460-462.

Mwaba, P., Mwansa, J., Chintu, C., Pobee, J., Scarborough, M., Portsmouth, S., and Zumla, A. (2001). Clinical presentation, natural history, and cumulative death rates of 230 adults with primary cryptococcal meningitis in Zambian AIDS